

DITERPENOIDS FROM *RABDOSIA JAPONICA*

XIANJUN MENG, QIGUANG WANG* and YAOZU CHEN†

Department of Chemistry, Lanzhou University, Lanzhou, P. R. China, *The Instrumental Analysis and Research Center of Lanzhou University, Lanzhou, P. R. China

(Received in revised form 29 July 1988)

Key Word Index—*Rabdosia japonica*; Labiatae, structural determination; diterpenoids, glaucocalactone; X-ray diffraction

Abstract—From the leaves of *Rabdosia japonica*, a new *ent*-kaurene lactone type diterpenoid, glaucocalactone, along with β -sitosterol, oridonin and rosthornin A were isolated. The structure of glaucocalactone was elucidated on the basis of spectral data and X-ray crystallographic studies.

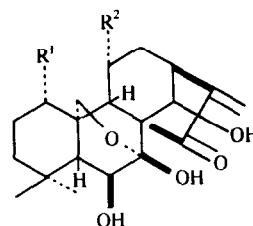
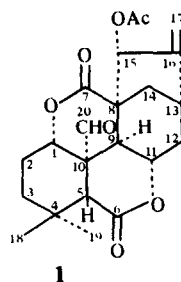
INTRODUCTION

The plants of *Rabdosia* species have been shown to contain some *ent*-kaurene type diterpenoids with cytotoxic activities [1–4]. In a continuation of our research work on diterpenoids, we have recently investigated *Rabdosia japonica* var. *glaucocalyx* collected from the southern Gansu Province of China. From the ether extract of the leaves, four crystalline compounds were isolated. One of the compounds is an *ent*-kaurene type lactone with a novel skeleton which we have named glaucocalactone (1), and the structure of which was elucidated as 1 on the basis of spectroscopic evidence and X-ray crystallographic studies. The other three compounds were identified as oridonin (2) [5, 6], rosthornin A (3) [7] and β -sitosterol. None of these compounds has been found in this species previously.

RESULTS AND DISCUSSION

Glaucocalactone (1), mp 318–320°, was isolated as colourless prisms. The mass spectral data and elemental analysis suggested its formula as $C_{22}H_{26}O_7$. The IR absorptions of 1 showed the presence of δ -lactone groups (1760, 1740 cm^{-1}), an aldehyde group (2750, 1720 cm^{-1}), a double bond (1660 cm^{-1}), and an acetoxy group (1725, 1240, 1040 cm^{-1}). The 400 MHz 1H NMR spectrum (Table 1) showed the signals of two tertiary methyl groups, two methine protons attached to oxygen-substituted carbon atoms, one methine proton attached to a carbon atom bearing an acetoxyl group, an aldehyde proton, an exomethylene and an acetoxy group. The ^{13}C NMR spectrum (Table 1) of 1 showed the presence of two lactonic carbon atoms, one aldehyde carbon atom, one acetoxy group along with five methylene carbon atoms, three methine carbon atoms, three oxygen-bearing methine carbon atoms, two methyl carbon atoms and four quaternary carbon atoms.

On comparison of the spectral data of 1 with those of closely related C-1 α , C-11 α and C-15 α hydroxyl-substituted



2 R¹=OH R²=H

3 R¹=H R²=OH

ted *ent*-kaurene compounds isolated from *Rabdosia* species [8–10], it appeared that glaucocalactone had a novel *ent*-kaurene lactone skeleton which is similar to that found in the *ent*-6,7-secokaurene diterpenoids [11].

The 1H NMR signal of H-15 at δ 5.92 (t, J = 2.5 Hz) is clear evidence for the existence of a 15 α acetoxy group while two signals at δ 4.84 (dd, J = 11.9, 4.2 Hz) and 5.00 (dt, J = 11.3, 8.7 Hz) could be assigned on H-1 β and H-11 β . This established that the two lactone linkages were C-1, O-1 α , C-7 and C-11, O-11 α , C-6. Moreover, the singlet signal at δ 10.00 indicated that the aldehyde group is attached to C-10.

The structure and stereochemistry of glaucocalactone were finally determined by X-ray crystallographic studies.

† Author to whom correspondence should be addressed

Table 1 ^1H and ^{13}C NMR data of compound 1

H		C	δ
1	4.84 <i>dd</i> (11.9, 4.2)*	1	75.50 (CH) [†]
2	2.34 <i>ddd</i> (13.0, 11.9, 5.2)	2	23.94 (CH ₂)
2	2.05 <i>dd</i> (13.0, 4.2)	3	38.47 (CH ₂)
3	1.54 <i>m</i> —	4	32.22 (C)
3	1.62 <i>m</i> —	5	49.52 (CH)
5	2.86 <i>s</i> —	6	169.38 (C) [‡]
9	2.43 <i>d</i> (11.3)	7	169.05 (C) [‡]
11	5.00 <i>dt</i> (11.3, 8.7)	8	49.72 (C) [§]
12	2.71 <i>ddd</i> (13.0, 8.7, 8.3)	9	39.49 (CH)
12	1.48 <i>dd</i> (13.0, 8.7)	10	48.23 (C) [§]
13	3.0 <i>dd</i> (8.3, 5.2)	11	68.62 (CH)
14	1.69 <i>dd</i> (12.5, 5.2)	12	36.24 (CH ₂)
14	2.23 <i>d</i> (12.5)	13	36.59 (CH)
15	5.92 <i>t</i> (2.5)	14	31.47 (CH ₂)
17	5.28 <i>dd</i> (2.5, 1.0)	15	80.77 (CH)
17	5.01 <i>br s</i>	16	152.19 (C)
20	10.00 <i>s</i>	17	111.50 (CH ₂)
Me-19	0.93 <i>s</i>	18	22.24 (Me)
Me-18	1.10 <i>s</i> —	19	29.43 (Me)
OAc	1.99 <i>s</i> —	20	202.26 (CH)
		21	171.55 (C)
		22	20.29 (Me)

The spectra were recorded at 400 MHz for ^1H and 100 MHz for ^{13}C NMR in DMSO- d_6 solution at 25 using TMS as internal standard.

* $J(\text{H}_2)$ in parentheses

[†] Assignment made with aid of DEPT

^{‡,§} Assignments may be interchanged

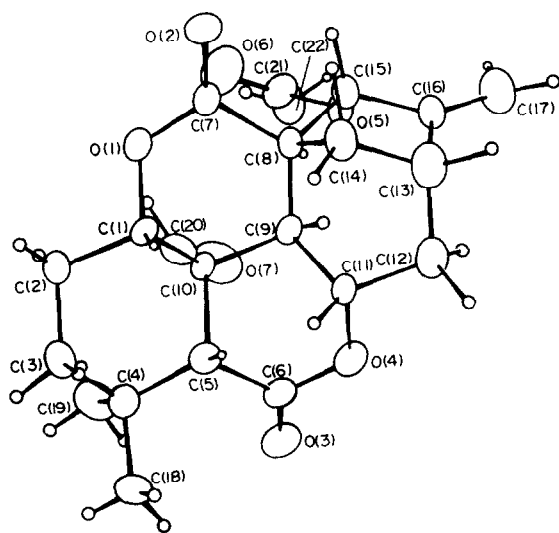


Fig. 1

The structure of glucocalactone was proved to be as shown in formula 1. The view of the molecule is given in Fig. 1. The crystallographic data and summary of intensity data collection and structure refinement of 1 are shown in Table 2. The positional parameters were given in Table 3. All the crystallographic details are deposited with the Cambridge Crystallographic Data Centre.

Table 2 Crystallographic data and summary of intensity data collection and structure refinement for compound 1

Molecular formula	$\text{C}_{22}\text{H}_{26}\text{O}_7$
M_r	402.45
Space group	$P2_12_12_1$
T, K	298
$a, \text{\AA}$	10.480 (2)
$b, \text{\AA}$	11.875 (2)
$c, \text{\AA}$	15.478 (3)
$V, \text{\AA}^3$	1926.2 (8)
Z	4
$d_{\text{calc}} \text{ g/cm}^3$	1.383
Crystal dimensions, mm	$0.6 \times 0.5 \times 0.1$
Radiation wavelength Cu K α , \AA	1.542
Crystal decay	< 1
μ, cm^{-1}	8.139
$F(000)$	856
Scan mode	$\omega/2\theta$
Scan width in $\omega, ^\circ$	$(0.70 + 0.15 \tan \theta)$
Aperture width, mm	$(1.5 + 0.4 \tan \theta)$
Aperture length, mm	4
Final acceptance limit	20° at 20 min
Maximum recording time, sec	60
Scan range, 2θ	2–55
No. of reflections collected	1662
No. of reflections observed (with $I > 3.0\sigma(I)$)	1640
$R = \Sigma F_o - F_c / \Sigma F_o $	0.070
R_w	0.080

Table 3 Positional parameters and equivalent isotropic temperature factors of non-hydrogen atoms for compound 1

Atom	$X (\times 10^4)$	$Y (\times 10^4)$	$Z (\times 10^4)$	$B_{\text{eq}} (\times 10)$
C(1)	11102 (7)	225 (6)	2101 (4)	27 (1)
C(2)	12073 (8)	543 (7)	1397 (5)	38 (1)
C(3)	13185 (8)	–303 (7)	1406 (5)	39 (2)
C(4)	13847 (7)	–474 (6)	2287 (5)	31 (2)
C(5)	12769 (7)	–734 (6)	2935 (4)	26 (1)
C(6)	13132 (8)	–1104 (6)	3846 (5)	33 (2)
C(7)	9088 (7)	909 (6)	2632 (4)	27 (1)
C(8)	9306 (7)	166 (6)	3439 (4)	25 (1)
C(9)	10690 (7)	–151 (5)	3683 (4)	23 (1)
C(10)	11703 (7)	212 (6)	3001 (4)	23 (1)
C(11)	10856 (8)	–1420 (6)	3896 (4)	29 (1)
C(12)	9872 (9)	–1850 (6)	4538 (5)	40 (2)
C(13)	8524 (8)	–1382 (7)	4304 (5)	40 (2)
C(14)	8475 (8)	–944 (6)	3365 (5)	35 (2)
C(15)	8600 (7)	715 (6)	4232 (4)	27 (1)
C(16)	8268 (7)	–288 (7)	4801 (5)	34 (2)
C(17)	7830 (9)	–211 (8)	5595 (5)	46 (2)
C(18)	14688 (8)	–1555 (7)	2222 (6)	43 (2)
C(19)	14687 (8)	536 (7)	2504 (6)	46 (2)
C(20)	12226 (7)	1360 (7)	3264 (5)	35 (2)
C(21)	9734 (9)	2460 (7)	4419 (5)	38 (2)
C(22)	10395 (9)	3173 (7)	5066 (6)	45 (2)
O(1)	10068 (5)	1044 (4)	2076 (3)	33 (1)
O(2)	8093 (5)	1348 (5)	2477 (3)	39 (1)
O(3)	14151 (6)	–1073 (5)	4171 (4)	49 (1)
O(4)	12105 (5)	–1529 (4)	4294 (3)	37 (1)
O(5)	9375 (5)	1480 (4)	4749 (3)	33 (1)
O(6)	9607 (8)	2669 (5)	3663 (4)	66 (2)
O(7)	12594 (6)	1564 (5)	3977 (4)	55 (1)

EXPERIMENTAL

Mps uncorr ^1H NMR 400 MHz with TMS as an internal standard ^{13}C NMR 100 MHz with TMS as an internal standard IR KBr pellets MS direct inlet, 70 eV X-ray CAD-4 diffractometer, computations: PDP 11 computer The plant material was collected by Mr Zhang Guoliang from the Liupan Mountain area of Gansu Province in July 1987 A voucher specimen has been deposited at the Herbarium of the Biology Department of Lanzhou University

Extraction and isolation The air-dried leaves of *R. japonica* var. *glaucocalyx* (2 l kg) were treated with Et_2O for 5 days at room temp After removal of Et_2O , 80 g of syrup remained The syrup was mixed with 100 g silica gel (100 mesh), extracted with petrol (30–60%) to remove lipids and pigments, and then subjected to CC on silica gel (200–300 mesh) eluting a petrol– Me_2CO gradient (from 5:1 to 2:1) Three fractions were collected The first fraction was rechromatographed on silica gel eluted with C_6H_6 –HOAc (7:1) to afford β -sitosterol (150 mg) Fraction 2 was subjected to further CC (CH_2Cl_2 – Me_2CO , 8:1) to afford glaucocalactone (1) (17 mg), oridonin (2) (175 mg) and orsthorin A(3) (90 mg) Each component was further purified by repeated recrystallization from an appropriate solvent

Glaucocalactone (1) Colourless prisms from CH_2Cl_2 , mp 318–320° (decomposed), $[\alpha]_D^{25} + 55^\circ$ (CHCl_3 , c 0.30), UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ) 200 (2.3), IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} 2750, 1760, 1740, 1725, 1720, 1660, 1240, 1060, 1040, EIMS m/z (rel int) 402 1682 $[\text{M}]^+$ (9), $(\text{C}_{22}\text{H}_{26}\text{O}_7)$ requires 402.1678, 360 $[\text{M} - \text{CH}_2\text{CO}]^+$ (55), 359 $[\text{M} - \text{MeCO}]^+$ (100), 331 $[\text{M} - \text{CH}_2\text{CO} - \text{CHO}]^+$ (12), 296 (13), 268 (12), 161 (28), 91 (25), 43 (95), ^1H and ^{13}C NMR Table 1 (Found C, 65.62, H, 6.50 $\text{C}_{22}\text{H}_{26}\text{O}_7$ requires C, 65.66, H, 6.51%)

Oridonin (2) Colourless needles from MeOH, mp 248–249°, $[\alpha]_D^{25} - 43.5^\circ$ (pyridine, c 0.80), UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ) 237 (3.94), IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} 3400–3200, 1700, 1640, 1095, 1080, 1060, EIMS m/z (rel int) 364 $[\text{M}]^+$ (8.6), 346 $[\text{M} - \text{H}_2\text{O}]^+$ (3.0), 328 $[\text{M} - 2\text{H}_2\text{O}]^+$ (2.6), 149 (7.9), ^1H NMR (400 MHz) δ (pyridine- d_5) 1.13, 1.29 (each 3H, Me-18, Me-19), 1.46 (1H, d , $J = 6.0$ Hz, H-5 β), 3.20 (1H, d , $J = 9.3$ Hz, H-13), 3.63 (1H, dd , $J = 6, 11$ Hz, H-6), 4.25 (1H, t , $J = 7.0$ Hz, H-1 β), 4.40, 4.77 (each 1H, AB-system, $J = 10$ Hz, H-20a, H-20b), 5.32 (1H, s , H-14 α), 5.50, 6.27 (each 1H, s , H-17a, H-17b), 6.00 (1H, $br s$, OH), 6.93 (1H, d , $J = 11$ Hz, 6-OH) (Found C, 65.89, H, 7.70 Calc for $\text{C}_{20}\text{H}_{28}\text{O}_6$, C, 65.92, H, 7.74%)

Rosthorin A(3) Colourless needles from MeOH, mp 257–259°, $[\alpha]_D^{25} - 79^\circ$ (pyridine, c 0.60), UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ) 236 (3.90), IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} 3470–3210, 1710, 1640, 1052, EIMS m/z (rel int)

364 $[\text{M}]^+$ (100), 346 $[\text{M} - \text{H}_2\text{O}]^+$ (8.5), 328 $[\text{M} - 2\text{H}_2\text{O}]^+$ (10), 318 (11), 285 (8), 215 (10), 151 (58), 85 (65), ^1H NMR (400 MHz) δ (pyridine- d_5) 1.10, 1.22 (each 3H, s , Me-18, Me-19), 1.60 (1H, d , $J = 6$ Hz, H-5 β), 1.94 (1H, dd , $J = 9, 13.8$ Hz, H-12 β), 2.18 (1H, d , $J = 8.7$ Hz, H-9 β), 3.05 (1H, ddd , $J = 9.9, 13.8$ Hz, H-12 α), 3.27 (1H, d , $J = 9$ Hz, H-13), 4.17, 4.31 (each 1H, AB-system, $J = 10$ Hz, H-20a, H-20b), 4.26 (1H, $br s$, H-6), 4.43 (1H, m , H-11 α), 5.20 (1H, s , H-14 α), 5.51, 6.25 (each 1H, s , H-17a, H-17b), ^{13}C NMR (100 MHz) δ (pyridine- d_5) 30.70 (C-1), 18.99 (C-2), 41.48 (C-3), 34.18 (C-4), 59.30 (C-5), 73.69 (C-6), 98.45 (C-7), 60.73 (C-8), 61.57 (C-9), 37.87 (C-10), 62.95 (C-11), 41.89 (C-12), 44.32 (C-13), 73.84 (C-14), 209.26 (C-15), 152.38 (C-16), 119.42 (C-17), 33.36 (C-18), 22.42 (C-19), 67.26 (C-20) (Found C, 65.98, H, 7.72 Calc for $\text{C}_{20}\text{H}_{28}\text{O}_6$, C, 65.92, H, 7.74%)

β -Sitosterol (4) Colourless needles from petrol, Mp 139–140°, $[\alpha]_D^{25} - 40^\circ$ (CHCl_3 , c 1.0), identical properties (mmp, IR) with an authentic sample of β -sitosterol.

Acknowledgements—We are grateful to the National Science Fund for financial support of this work Thanks are also due to the Instrumental analysis and Research Center of Lanzhou University for MS, UV and IR spectral measurements and elemental analysis

REFERENCES

1. Li, J. C., Liu, C. J., An, X. Z., Sun, H. D. and Lin, Z. W. (1984) *Acta Botan. Yunnan* **6**, 453
2. Wang, Z. Q., Wang, X. R., Dong, I. G., Xue, Z. W. and Wang, X. W. (1984) *Acta Chim. Sinica* **45**, 871
3. Fujita, E., Nagao, Y., Kohno, T., Matuda, M. and Ozaki, M. (1981) *Chem. Pharm. Bull.* **29**, 3208
4. Cheng, P. Y., Xiu, M. I., Lin, Y. L. and Shi, I. C. (1982) *Acta Pharmaceut. Sinica* **17**, 917
5. Fujita, E., Fujita, T., Katayama, H. and Shibuya, M. (1970) *J. Chem. Soc. (C)* 1674
6. Fujita, E. and Taoka, M. (1972) *Chem. Pharm. Bull.* **20**, 1752
7. Li, G. Y. and Wang, Y. L. (1984) *Acta Pharmaceut. Sinica* **19**, 590
8. Fujita, E., Fujita, T., Taoka, M., Katayama, H. and Shibuya, M. (1973) *Chem. Pharm. Bull.* **21**, 1357
9. Ochi, M., Okamura, M., Kotsuki, H., Miura, I., Kubo, I. and Kubota, T. (1981) *Bull. Chem. Soc. Jpn* **54**, 2786
10. Xiu, G. Q. (1985) *Acta Chim. Sinica* **43**, 35
11. Fujita, E. and Node, M. (1984) in *Progress in the Chemistry of Organic Natural Products* **46**, p. 80 Springer, Wien